



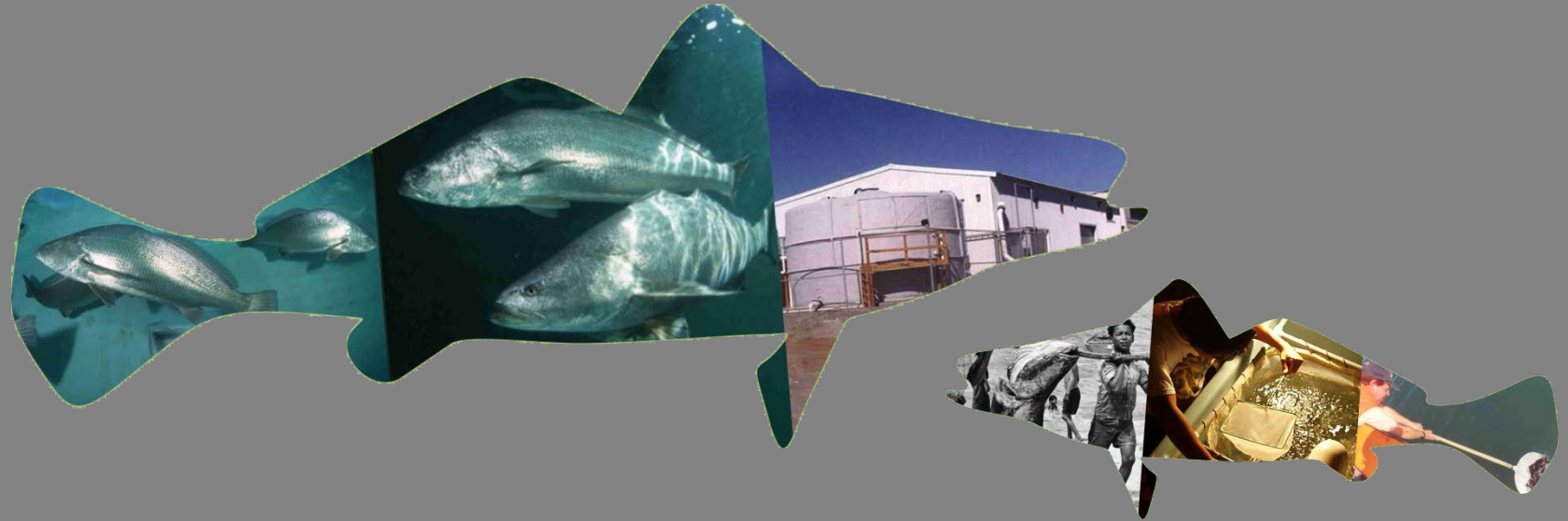
# Genetic traceability of wild and captive-reared totoaba stocks: A tool for population recovery assessment and law enforcement

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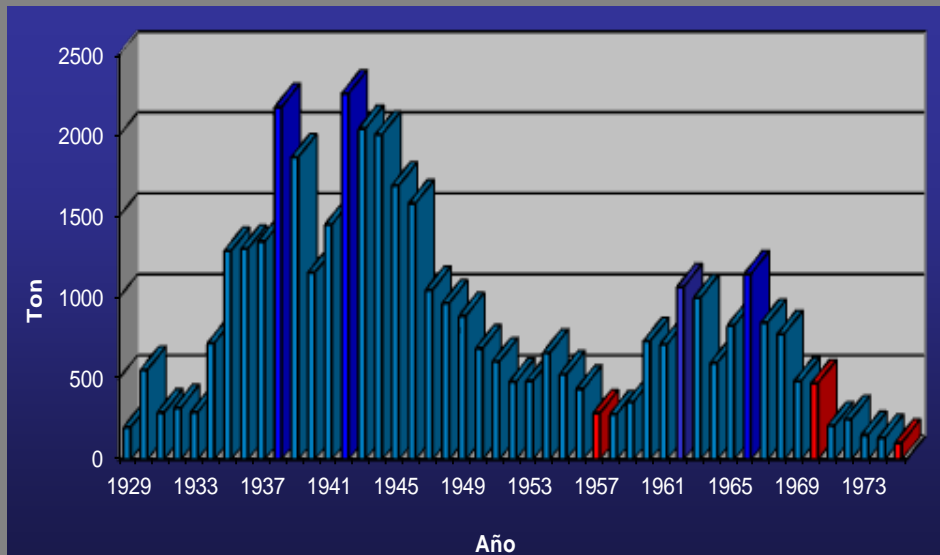
# Background



*Totoaba macdonaldi*

## Data:

- Max Age ( $L_{\infty}$ ) 50 years.
- Max Rep. Length 2.30 m.
- Size at First Year 50 cm.
- Weight at First Year 1-3 kg.
- Max Weight 150 kg



## Habitat:

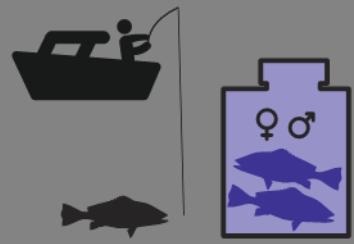
- Endemic to the Gulf of California
- Distribution / Migration Pattern



## Importance: Historic/Ecologic/Economic

- Commercial and Sport Fishery
- Historical Value due to Swim Bladder
  - NOM-059-ECOL-2001
  - CITES Appendix I
- Meat firm white low fat content.





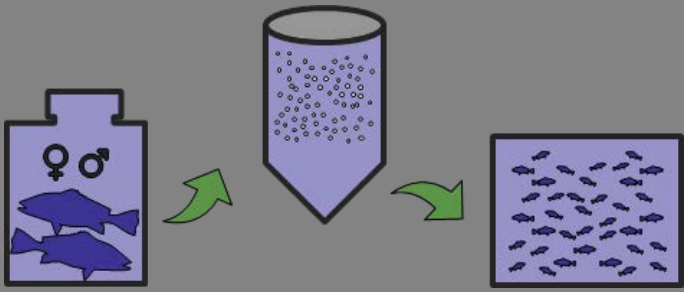
# Brood Stock acquisition and maintenance



- Four collections over the past 20 yrs. (60 wild BS)
- Improvements in decompression procedure during capture, transport and acclimation results in 50-65% survival.
- Maturation in captivity with an optimal 10 years period of egg production once they are 8 y.o.
- Observed fecundity ~ 1 million eggs for a 10 kg female and an additional million for every kg of weight gain.
- Fertile eggs are produced through manipulation of environment and assisted by hormonal treatment.



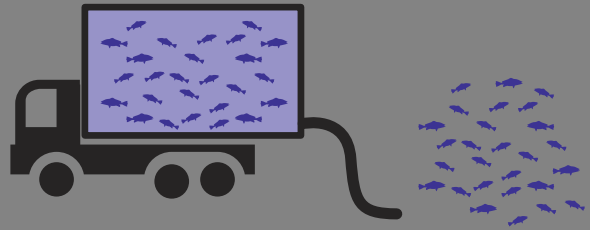
# Captive Breeding Biotechnology



- Selective breeding procedures: Pedigree
- Improved protocols for larvae to juvenile rearing:
  - ✓ T°C / Light / Stocking densities / Feeding with species specific nutritional requirements, etc.
  - ✓ Scaled up to produce 50,000 juveniles from each million eggs (5% survival).
  - ✓ Cannibalism / Size disparity / Deformities
- Base Unit (Lab) with overall costs estimates based on real production from BS capture to fingerlings.
- Current installed capacity
  - ✓ 16 Wild-BS (1♀:1♂) , 38 F1-BS (14♀:10♂:14?)
  - ✓ Fertile egg production up to 35 millions
  - ✓ Larval rearing up to 1.5 million / run (max. 3 runs)
  - ✓ Limited space for weaning and grow out.







# Experimental Releases

- Eight experimental releases into wild stock starting in 1997: Nearly 23,000 individuals.
- Physical / Genetic tagging:
  - Evaluation of stock enhancement feasibility
  - First genetically tagged recaptures in 2014
- Awareness: Community participation



**¿Ayudarías a recuperar la Totoaba?**

DESDE HACE ALGUNOS AÑOS SE HA TRABAJADO INTENSAMENTE...

En el Laboratorio de Biotecnología en Piscicultura de la Universidad Autónoma de Baja California, desde 1994 se cuenta con reproductores de totoaba y en 1996 se logró la primera reproducción en cautiverio. Recientemente inició el programa de repoblamiento con la intención de recuperar este valioso recurso. Los juveniles de totoaba que se están liberando cuentan con una marca que los distingue de los del medio natural.

**¿CÓMO LOGRAREMOS LA RECUPERACIÓN DE ESTE RECURSO?**

La idea consiste en liberar miles de juveniles de totoaba, para que en un futuro se reproduzcan con las que aún viven en el golfo y de esta manera la población se recupere.

**SI LLEGARA A CAER EN TUS REDES UNA TOTOABA MARCADA**

Con tus reportes podremos evaluar el crecimiento y la sobrevivencia de los juveniles que se están liberando. Al encontrar una totoaba marcada es importante que conserves su cabeza. Puedes entregarla en las oficinas de la SEMARNAT o comunicarte a la FACULTAD DE CIENCIAS MARINAS al teléfono (617) 4570 extensión 121 o envíenos al correo electrónico [totoaba@biotecna.uabc.mx](mailto:totoaba@biotecna.uabc.mx). Si es posible registra el tamaño, el peso y la zona donde encontraste la totoaba. No habrá averiguación alguna.

La marca utilizada es un tatuaje de color naranja fluorescente colado en la lengua, del tamaño de un grano de arroz. Esta marca permanece en los siguientes por el resto de su vida sin causarles daño alguno.

**Salvemos a la Totoaba**

Para que exista como en los viejos tiempos y la aprovechemos racionalmente en los nuevos tiempos

La UABC está desarrollando el cultivo de Totoaba con el fin de repoblar el Golfo de California, pero requerimos de TU AYUDA.

Al caer una Totoaba en tus redes revisa la lengua, ya que las Totoabas nacidas en la Unidad de Biotecnología en Piscicultura (UBP) de la UABC se marcan en la lengua con un tatuaje azul semejante al tamaño de un grano de arroz.

Si presenta la marca, mide a la Totoaba y entrega su cabeza en las oficinas de SEMARNAT y notifiéalo al teléfono 01 (646) 174-45-70, ext. 121 o en el correo electrónico: [totoaba@uabc.mx](mailto:totoaba@uabc.mx).

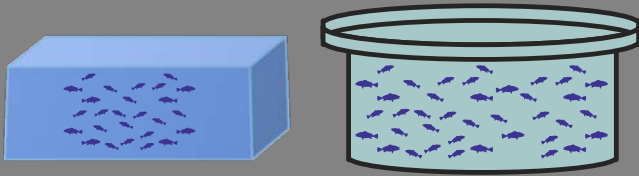
**RECUERDA QUE LA PESCA DE LA TOTOABA ESTÁ PROHIBIDA.**

Totoaba marcada en la lengua

**Salvemos a la Totoaba**

Para que exista como en los viejos tiempos y la aprovechemos racionalmente en los nuevos tiempos

La UABC está desarrollando el cultivo de Totoaba con el fin de repoblar el Golfo de California, pero requerimos de tu ayuda. Al caer una Totoaba en tus redes revisa la lengua, ya que las Totoabas nacidas en la Unidad de Biotecnología en Piscicultura de la UABC se marcan en la lengua con un tatuaje azul semejante al tamaño de un grano de arroz. Si presenta la marca, mide a la Totoaba y entrega su cabeza en las oficinas de la SEMARNAT y notifiéalo al teléfono 01(646) 174-45-70, ext. 121 ó en el correo electrónico: [totoaba@uabc.mx](mailto:totoaba@uabc.mx). Recuerda que la pesca de Totoaba está prohibida.



# Mariculture Development

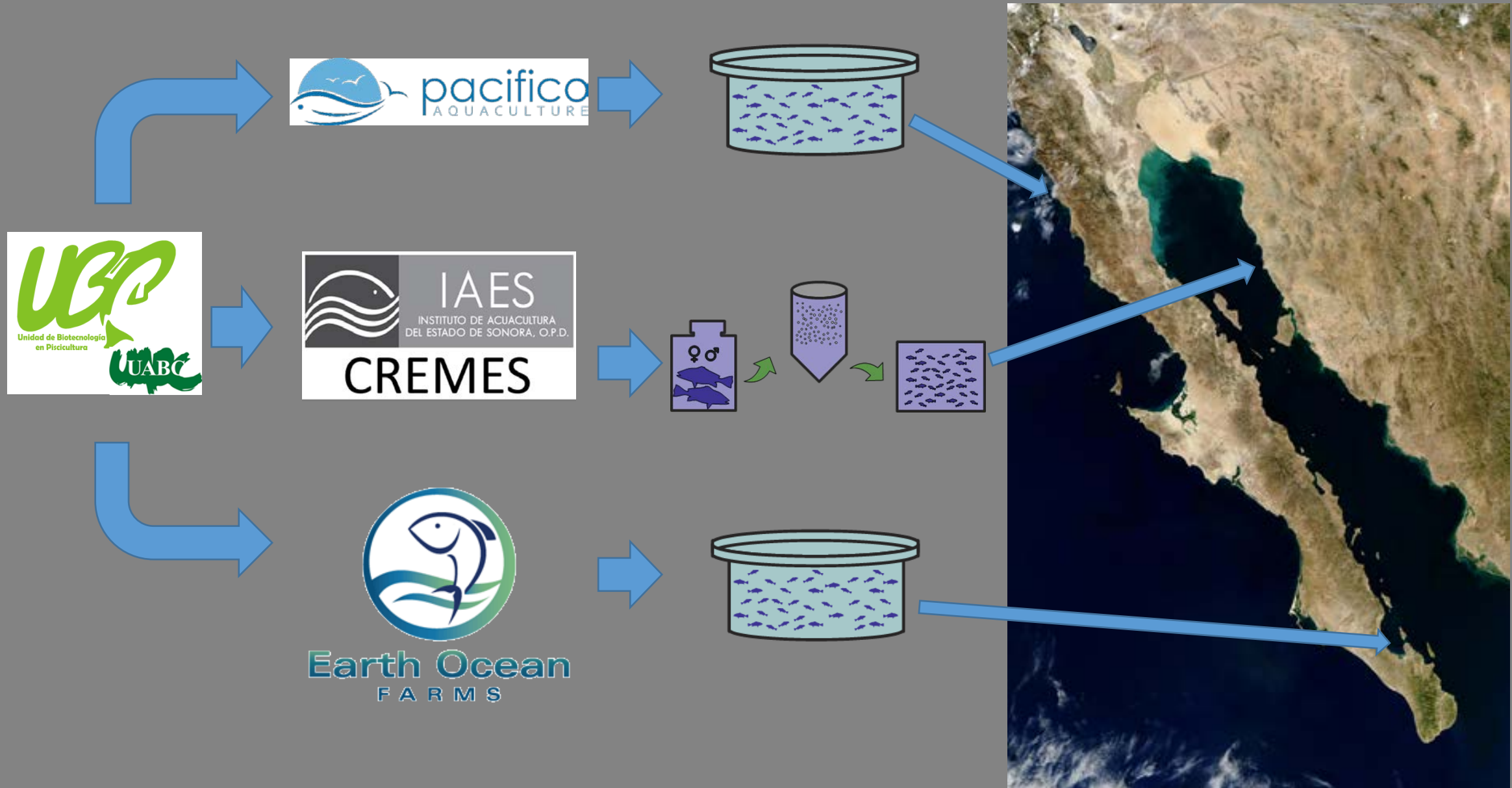


- Land-based grow out trial \*
  - ✓ PEZCO / PROVIPSA
  - ✓ 2010 First authorization to sell captive-bred totoaba.
  - ✓ Selection of F1 generation animals for aquaculture-oriented brood stock.
  - ✓ UABC: First captive breeding unit registered as an Environmental Management Unit (UMA).
- Marine farms development \*
  - Delly / Pacifico / EOF: Lots from UABC's UMA
  - Surface and submerged cages
  - Current development of cost-effective diet for grow out

\* All productive and commercial activities under NOM-059 regulation (SEMARNAT-DGVS).



# Current Totoaba UMAs





# Genetic Markers: Objectives

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## Selective captive breeding program:

- ✓ Constrain inbreeding and pedigree selection
- ✓ Evaluate performance of wild and captive reared broodstock

## Genetic forensics:

- ✓ Species ID for detection of illegal products
- ✓ Individual traceability to regulate commercialization
- ✓ Ensure transparency in law enforcement

## Population dynamics:

- ✓ Estimate current and historical population effective size
- ✓ Demographic trend inference

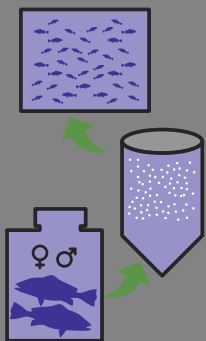




# Captive Breeding Program

## Genetic diversity and relatedness:

- ✓ High nuclear and mitochondrial diversity ( $H_o = 0.71$ ;  $H = 0.91$ )
- ✓ Very low relatedness within broodstock ( $R = 0.02$ )

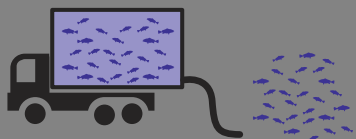


## Parental assignment:

- ✓ 100% assignment using only 12 out of 24 tested STRs
- ✓ Low probability of identity among full sibs (1 : 1,500 millions)

## Experimental releases to wild stock:

- ✓ Nearly 23,000 color-tagged fingerlings (90% genetic tag)
- ✓ Two seized bladders tracked back to captive breeding







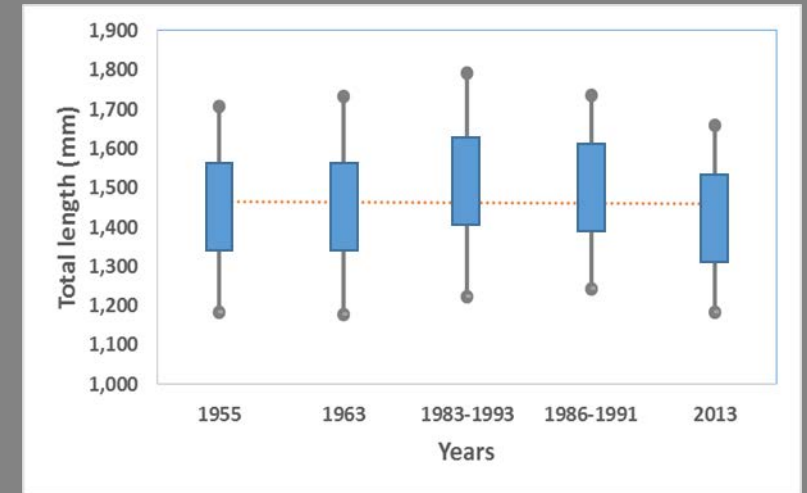
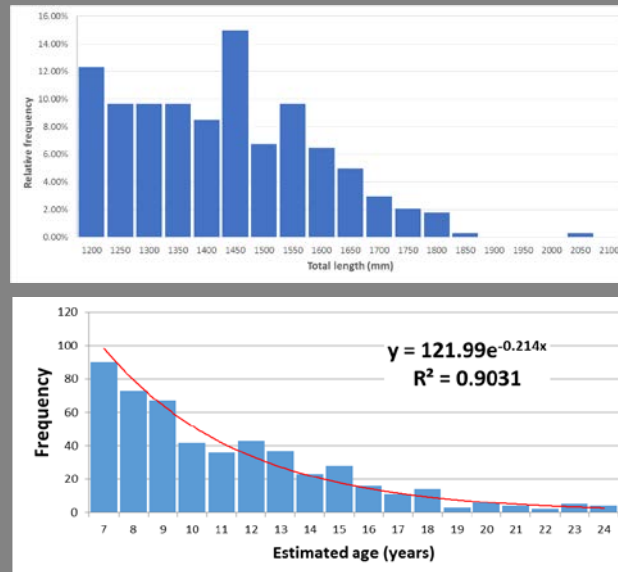
# Wild stock dynamics

## Morphometric data from seizures: No sign of stock depletion

- ✓ No significant shifts in size and age structure in the last 60 years
- ✓ Age of first reproduction, individual growth rate and adult mortality are also the same than in early 50's. All above suggest a stable stock.



USFWS / PROFEPA / PGR



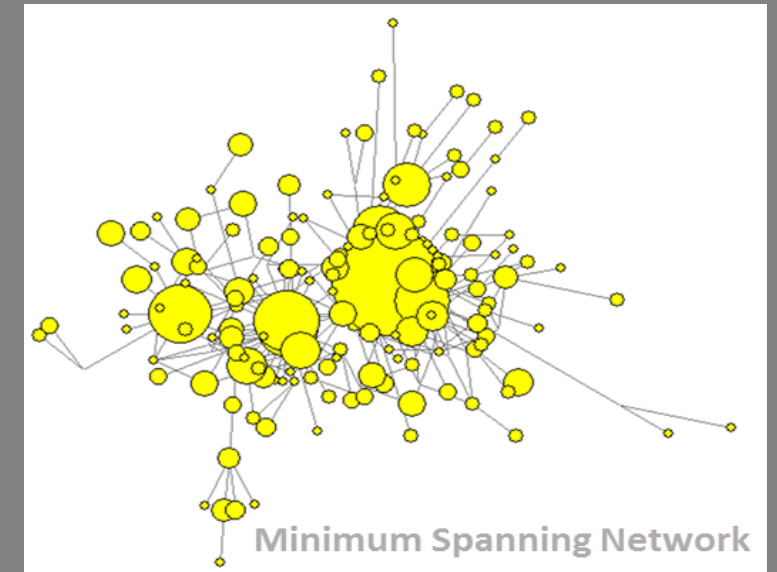
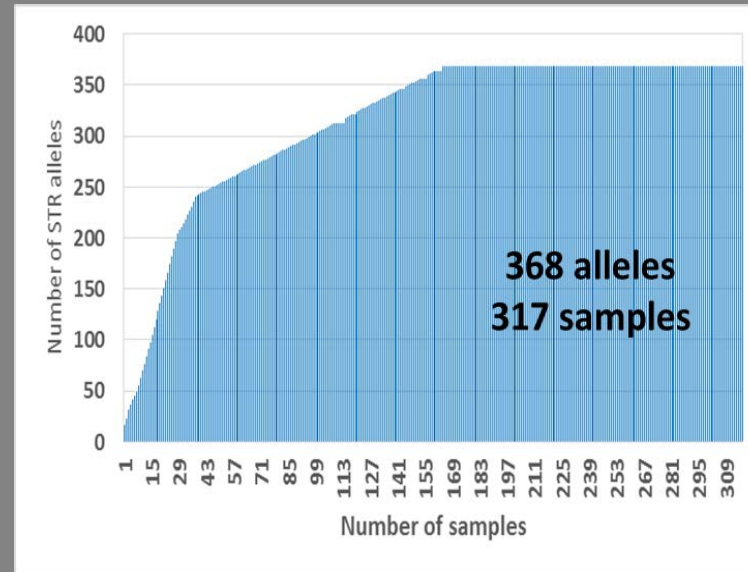
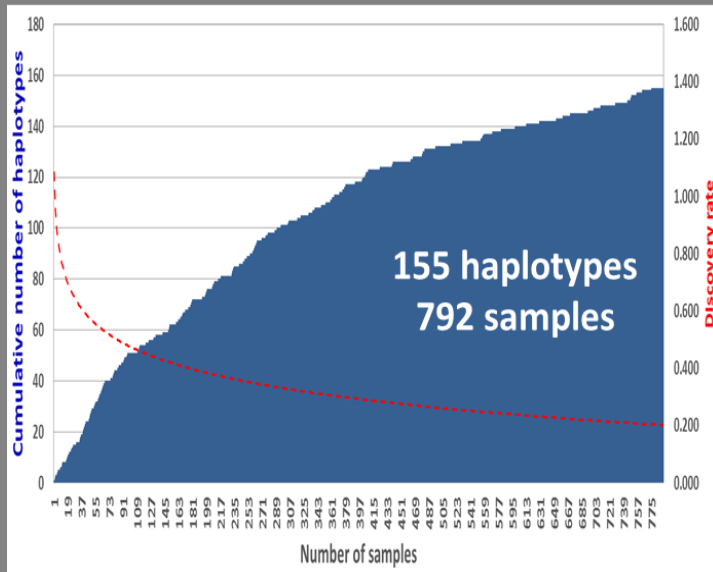




# Wild stock dynamics

## Genetic diversity and demographic history:

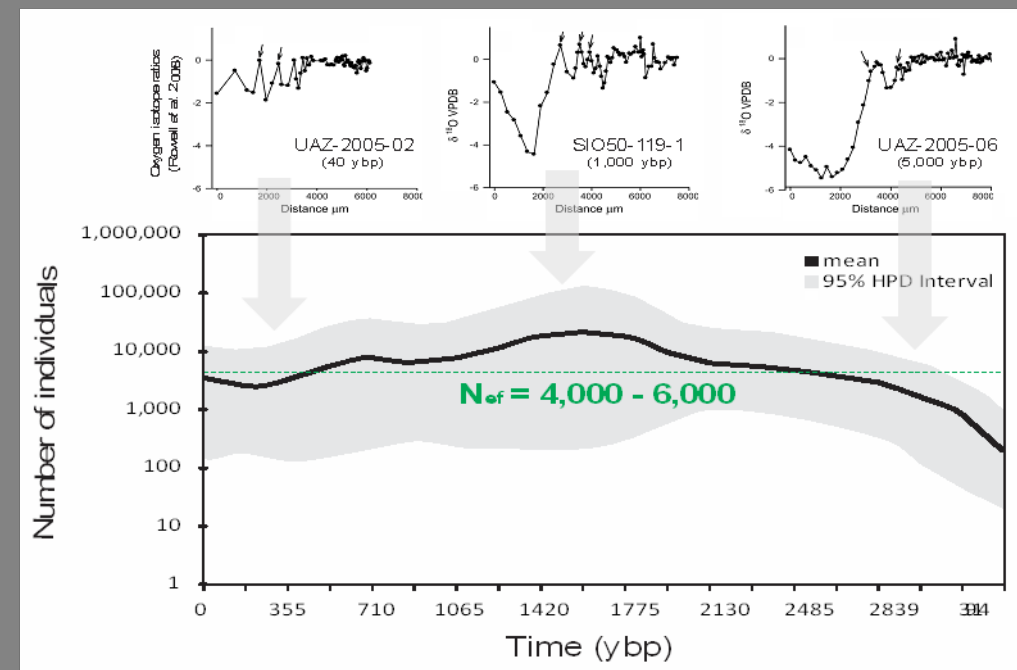
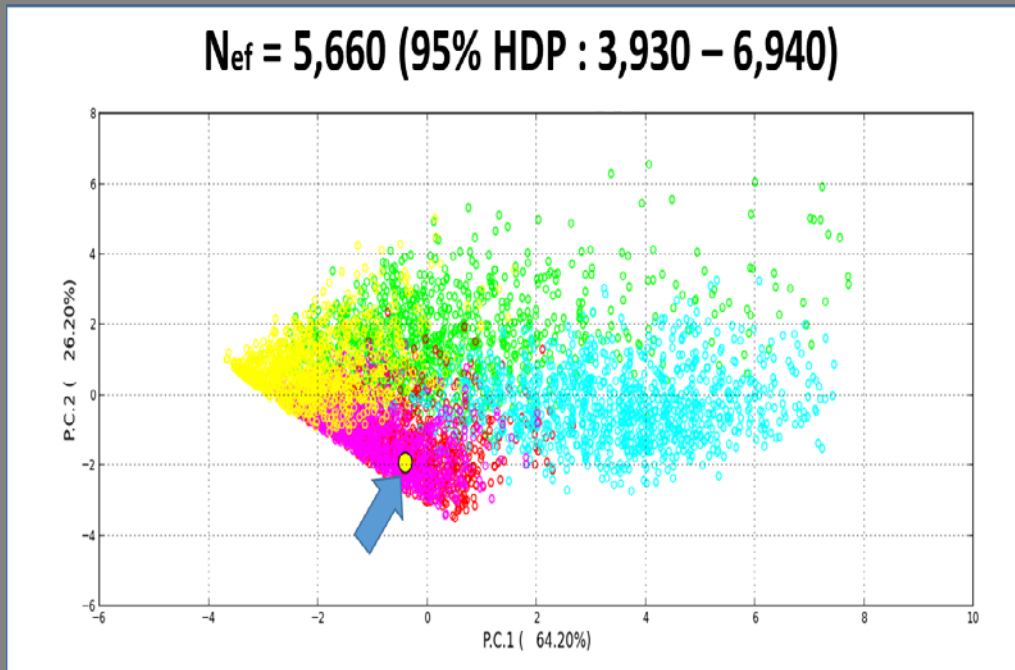
- ✓ High genetic diversity is not congruent with assumed fishery collapse.
- ✓ Demographic tests do not support a recent severe bottleneck.





## Effective Population Size ( $N_e$ )

- ✓ Estimated  $N_e$  supports a stable demography and ensures long term conservation of genetic diversity (Enríquez *et al.* 2008, Valenzuela-Quiñones *et al.* 2014, UABC 2015).
- ✓ Major demographic events depicted from genetic data did not occurred within the timeframe of anthropogenic impact to totoaba population (Valenzuela-Quiñones *et al.* 2014, UABC 2015).

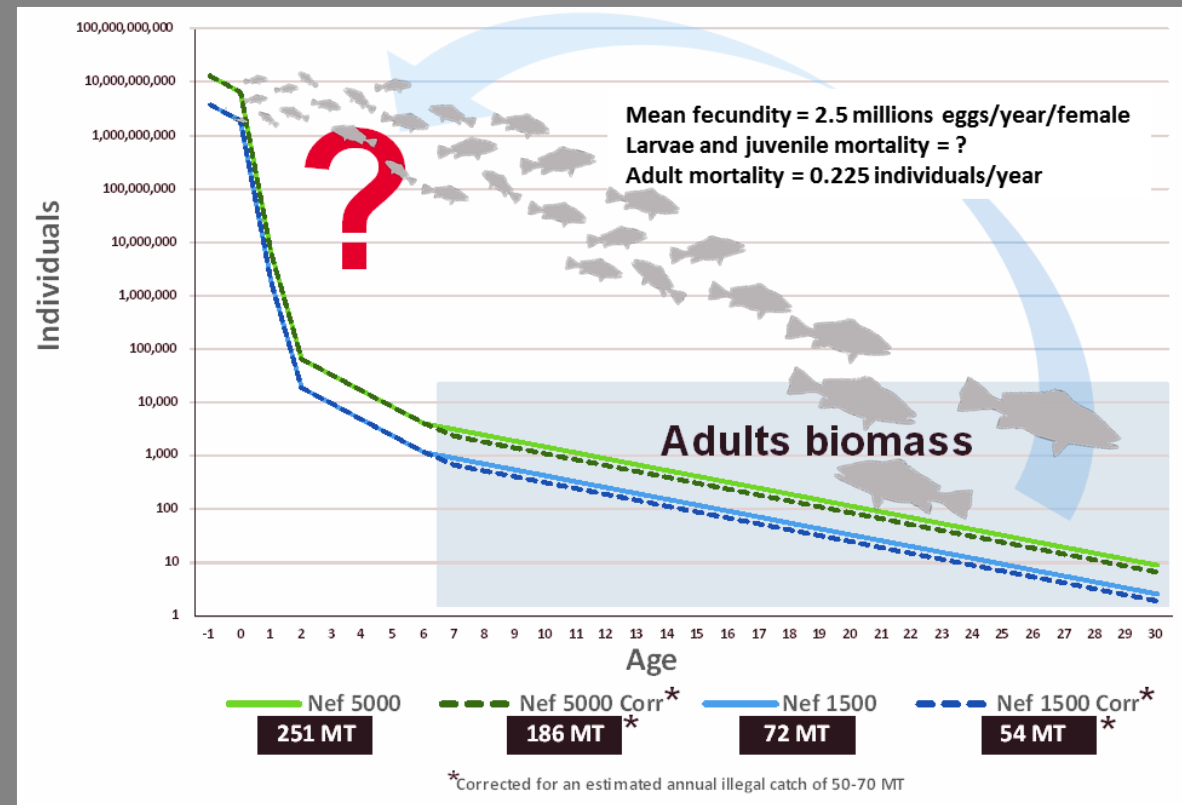
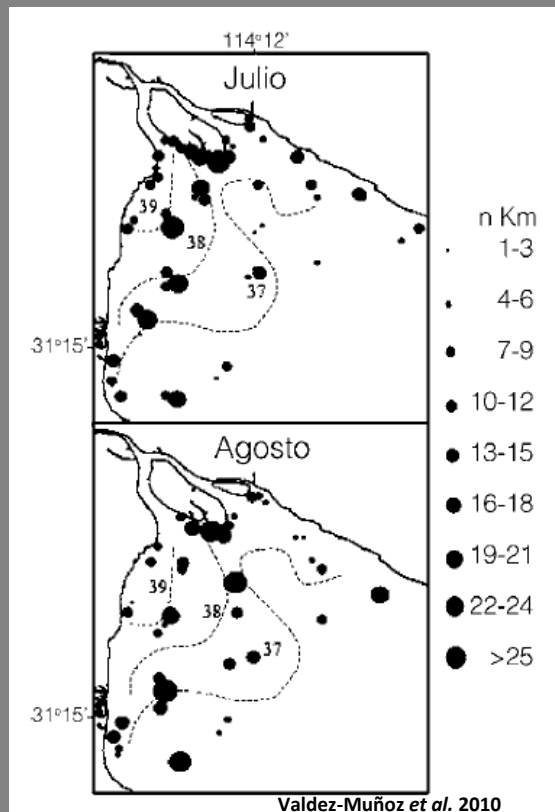




# Wild stock dynamics

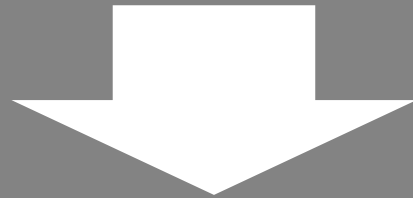
## Demographic trend simulations:

- ✓ Simulations based on effective population size estimates and current knowledge on totoaba life history suggest juvenile mortality as the main issue in stock recovery.



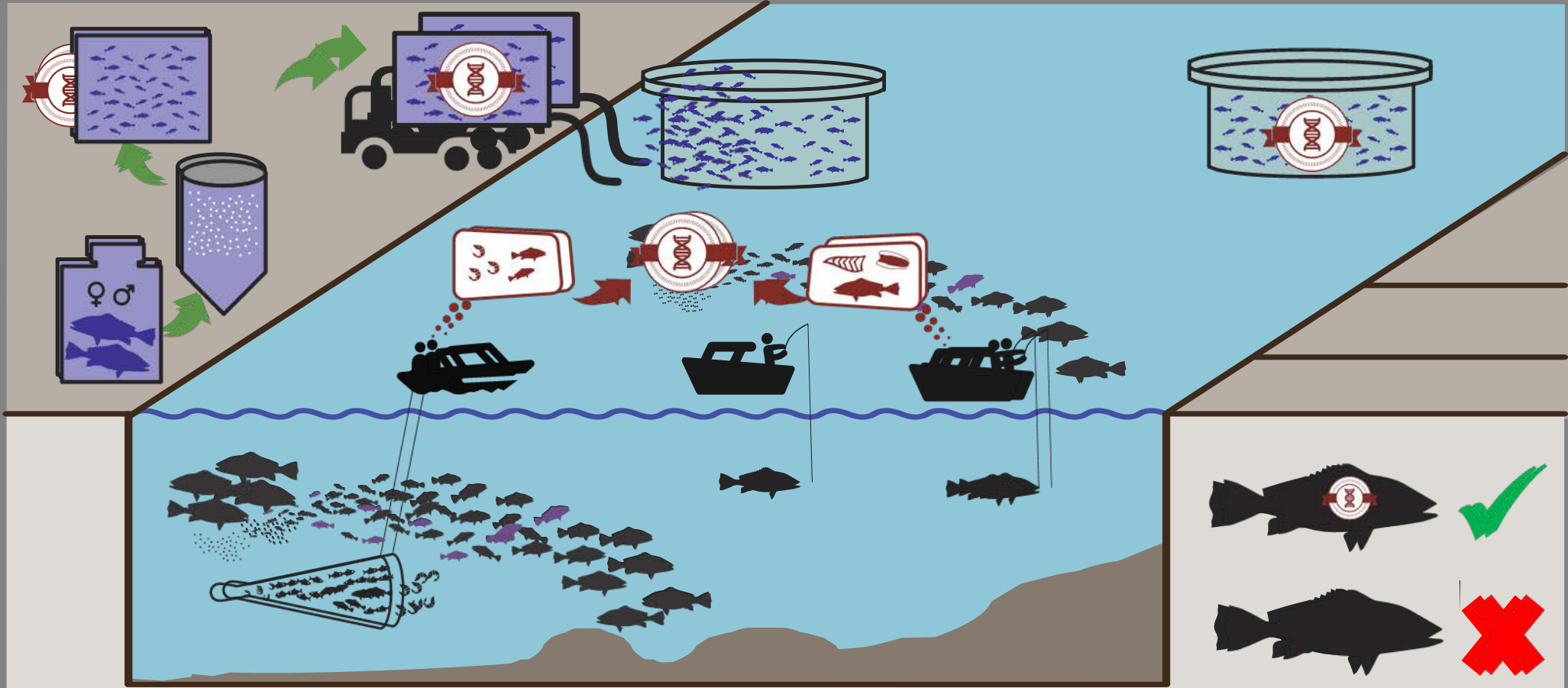


- ✓ Captive breeding technology is fully developed and has spread.
- ✓ Traceability can be guaranteed, down to individual level.
- ✓ Current available data suggest totoaba wild-stock is not critically endangered.
- ✓ First two recaptures of captive-reared totoaba: Survival to adult stage.



## Stock Enhancement Feasibility

# Genetically Monitored Stock Enhancement



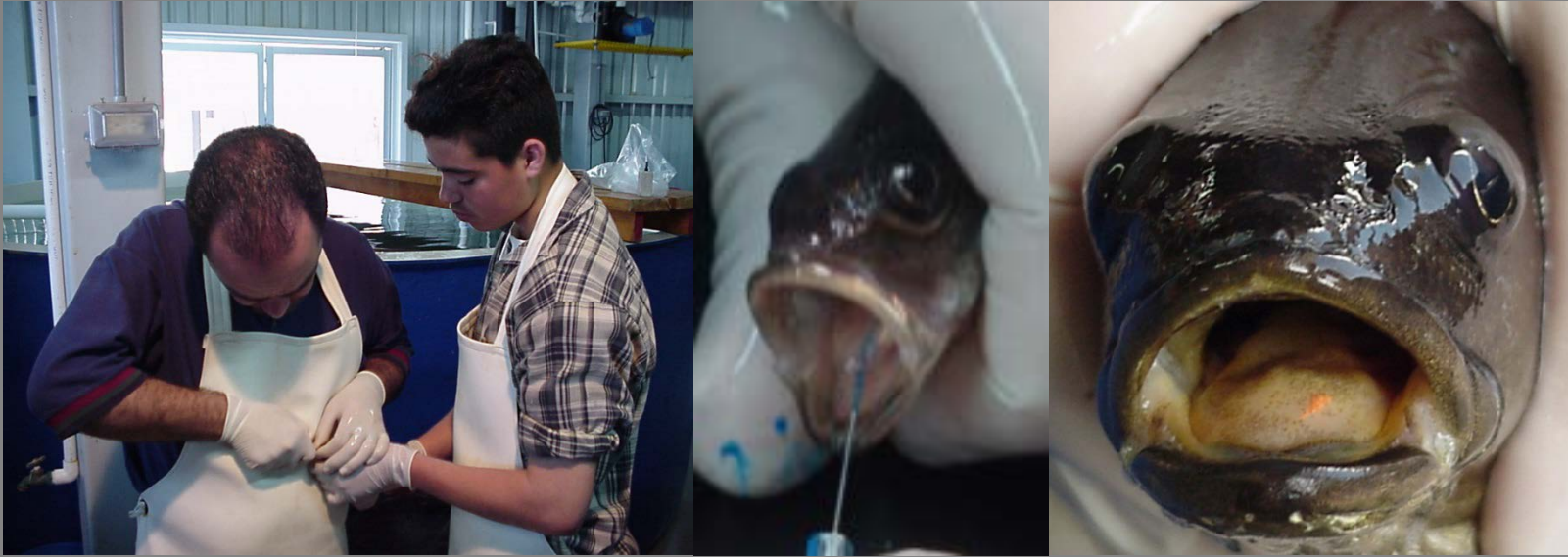
➡ **Reduction in stock** → **Long-term wild-stock genetic monitoring**

# Requested Specific Outcomes

- Collaborate to complete the standardization, optimization and validation of a high-resolution and low-cost genetic markers set for genetic traceability of totoaba captive-reared and wild stocks.
- Improve and scale-up, with support from the corresponding Governmental Agencies, current breeding and rearing capabilities at Environmental Management Units (UMAs) in order to properly start a stock enhancement pilot program.
- Set an agreement with the Mexican Federal Governmental Agencies to establish a mandatory genetic traceability program. Building up a traceability data base from all available sources (captive breeders, fingerlings by-catch, seizures, scientific collections and recreational catch, if any) is critical for law enforcement and long-term genetic monitoring wild stock.



Species	IUCN	N	n <sub>H</sub>	$\pi_n$	H	Annual Catch	Reference
<i>Atractoscion aequidens</i>	NE	104	32	0.005	0.85		Henriques et al. 2014
<i>Atractoscion aequidens</i>	NA	40	19	0.008	0.90		Henriques et al. 2014
<i>Atractoscion nobilis</i>	LC	59	32	0.010	0.96	1,500 MT	Ríos (2010), SAGARPA (2013)
<i>Collichthys lucidus</i> *	NA	151	55	0.034	0.89	320,000 MT	SNFRI (2001), Song et al. (2013)
<i>Cynoscion acoupa</i>	LC	297	83	0.003	0.89	15,000 MT	Fundación PROZEE (2006), Rodrigues et al. (2008)
<i>Cynoscion nebulosus</i> *	NA	280	60	0.024	0.85	4,500 MT	NMFS (2000), Anderson y Karel (2010)
<i>Larymichthys polyactis</i> *	NA	127	125	0.013	1.00	388,000 MT	FAO (2010), Wu et al. (2012)
<i>Nibea albiflora</i>	NA	65	37	0.008	0.97		Han et al. 2008a
<i>Pennahia argentata</i> *	NA	132	113	0.026	1.00		Han et al. 2008b
<i>Sciaenops ocellatus</i> *	NA	209	134	0.030	0.98	700 MT	Van Voorhees et al. (1992), Seyoum et al. (2000)
<i>Cynoscion reticulatus</i>	LC	42	31	0.016	0.99		Ríos 2012
<i>Cynoscion othonopterus</i>	V	92	23	0.004	0.69	2,600 MT	Rodriguez-Quiroz et al. (2010), Ríos (2012)
<i>Totoaba macdonaldi</i>	CE	74	42	0.013	0.98		Quezada 2009
<i>Totoaba macdonaldi</i>	CE	792	155	0.013	0.96	60 MT	UABC 2014



Year/Color	N	Current age
1997	200	17 y.o.
1999	600	15 y.o.
2000	1200	14 y.o.
* 2001	2100	13 y.o.
2002	1600	12 y.o.
* 2007	2000	7 y.o.
2012	2070	2 y.o.
2013	12000	1 y.o.



**22,970**

**individuals tagged and  
released to the wild**



Cohort	# Released	Current age
1997	200	17
1999	600	15
2000	1,200	14
2001	2,100	13
2002	1,600	12
2007	2,000	7
2012	2,070	2
2013	12,000	1



1 Recapture?  
UBP (18 STRs)



1 Recapture  
UBP (24 STRs)





Gimlet

Identification Calculator Input file Output file and Go! Help About

 **version 1.3.3** Kinship

"Exact" Kinship determination

Accept kinship with up to  incompatibilities (number of loci)

Missing alleles: ☒ treat as any allele Determine: ☒ 1 parent ☐ pair

☐ exclude locus ☐ verify kinship ☐ limit the kinship

☐ Use demographic data to confirm genetic kinship

In information file:

Choose file!

Years of Birth/Death  
+/-

☐ Age of first reproduction

Male	<input type="text"/>	year(s) +/-	<input type="text"/>
Female	<input type="text"/>	year(s) +/-	<input type="text"/>
Undet.	<input type="text"/>	year(s) +/-	<input type="text"/>

☐ Time between birth and conception

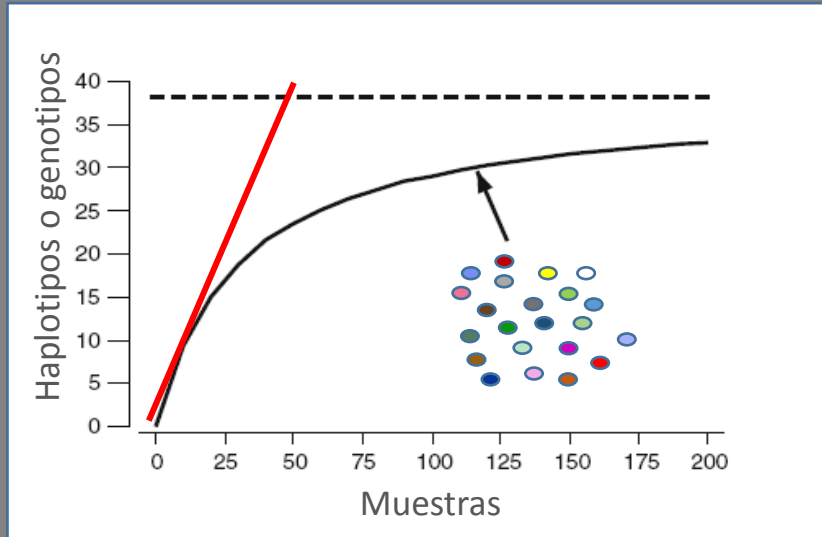
year(s)  
+/-

Parentage (2P) - Notepad

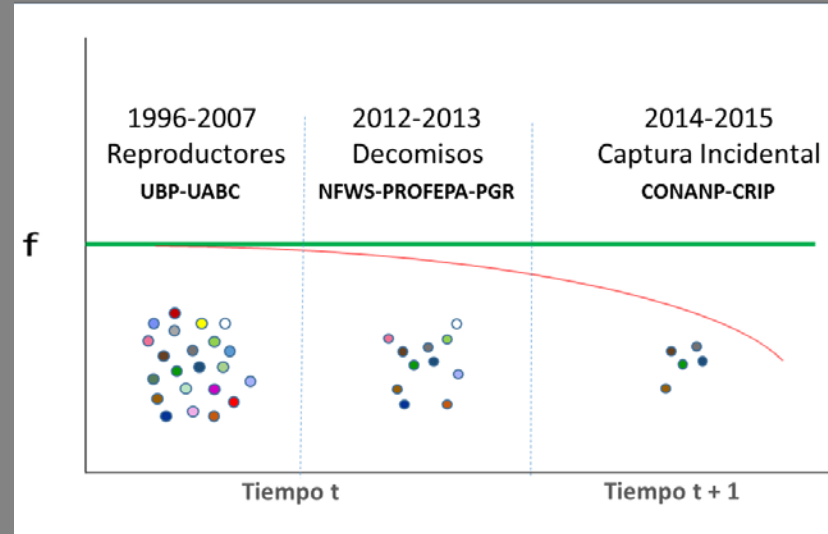
File Edit Format View Help

```
Gimlet Software v1.3.3 by N.VALIERE, 2003
Date: 10-08-2014 17:02:21
Maximum number of incompatibilities= 0
Results for the determination of the two parents
Potential pairs of parents
(Mother/Father when sex is available)
TA0322012      = parent
TA0462007      = parent
TA0472007      = parent
TA0482007      = parent
1PG1097        orphan ? :(
1PG1139        TA0322012/TA0752007 (*)
1PG7134        orphan ? :(
1PG7215        orphan ? :(
1PG7101        TA0482007/TA0672007 (*)
1PG7106        orphan ? :(
1PG7125        orphan ? :(
1PG7222        orphan ? :(
1PG7225        orphan ? :(
```

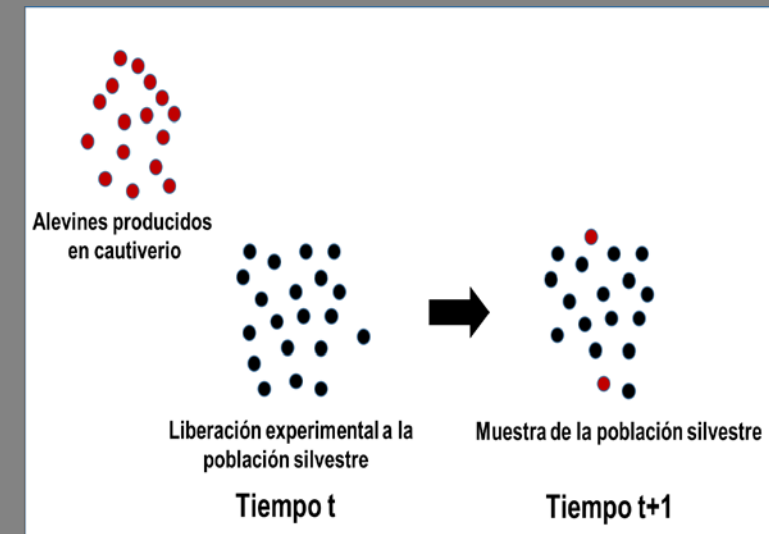
## Rarefaction (Discovery rate)



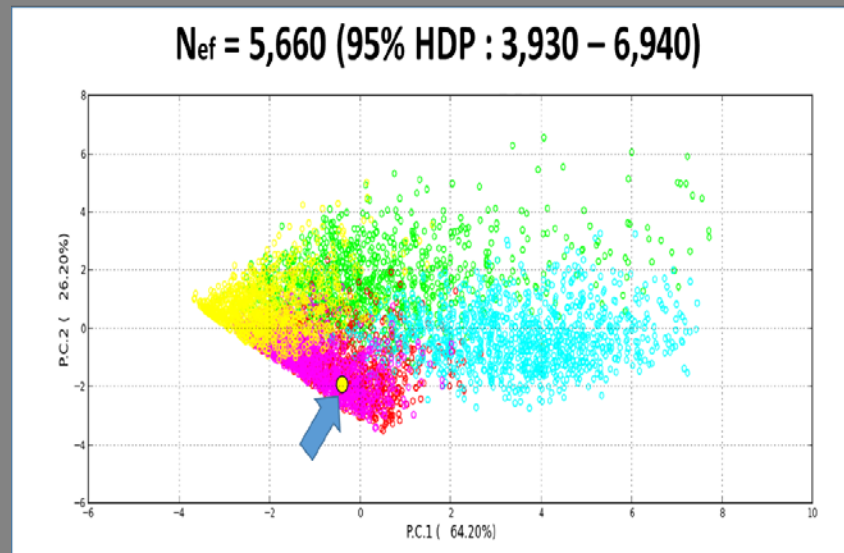
## Temporal Shifts in Allele Frequency



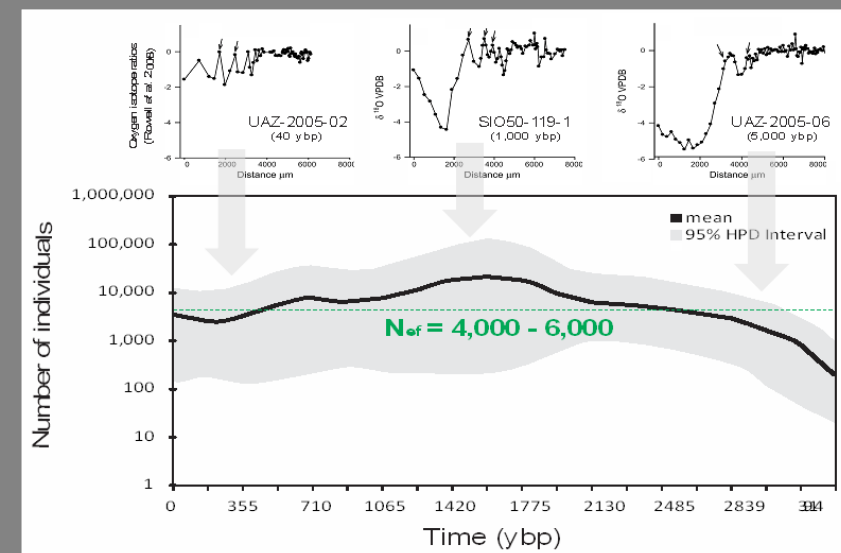
## Capture - Recapture



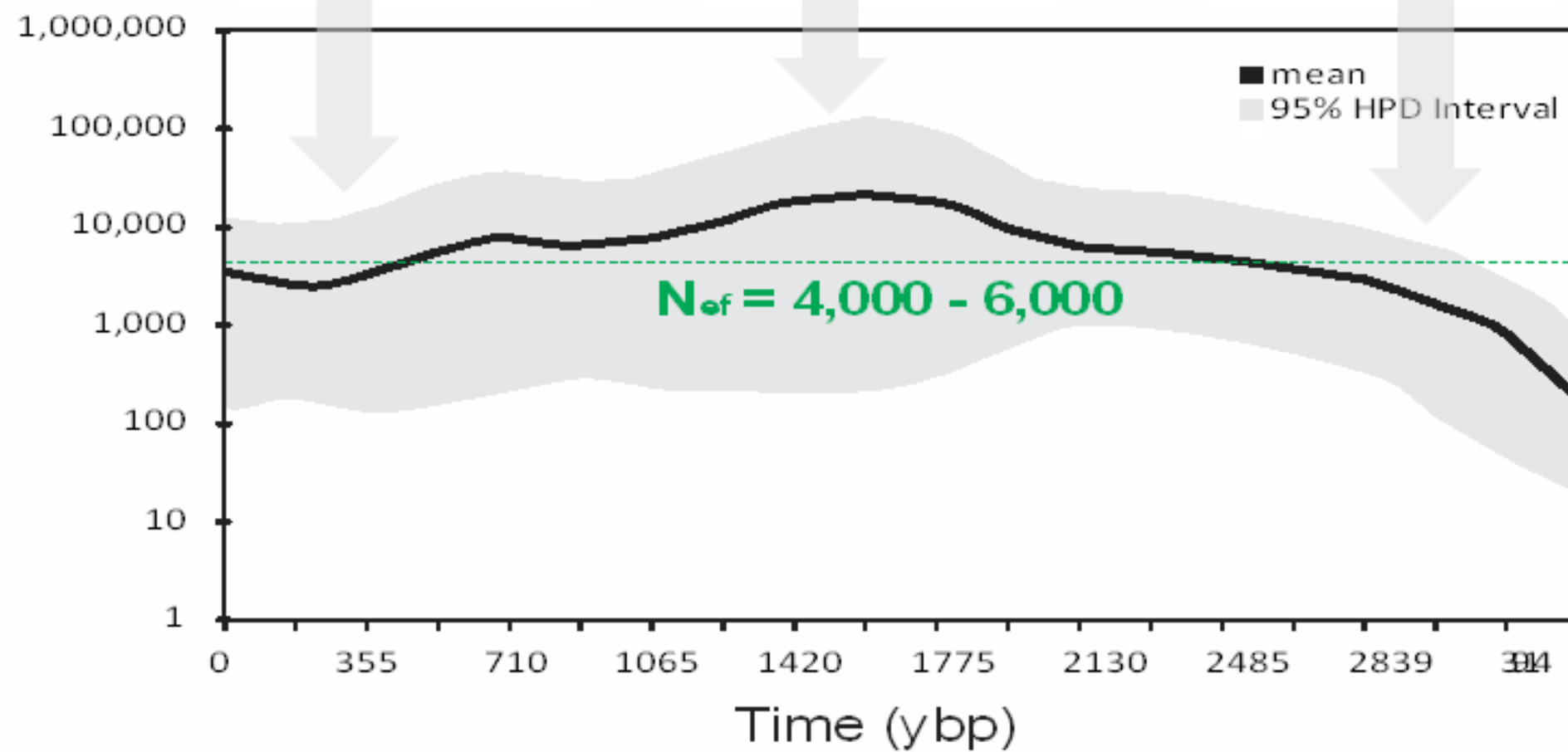
## Aproximate Bayesian Computation Past Inference



## Coalescent Bayesian Skyline Plot



Number of individuals



## Effective population size (breeding adults)

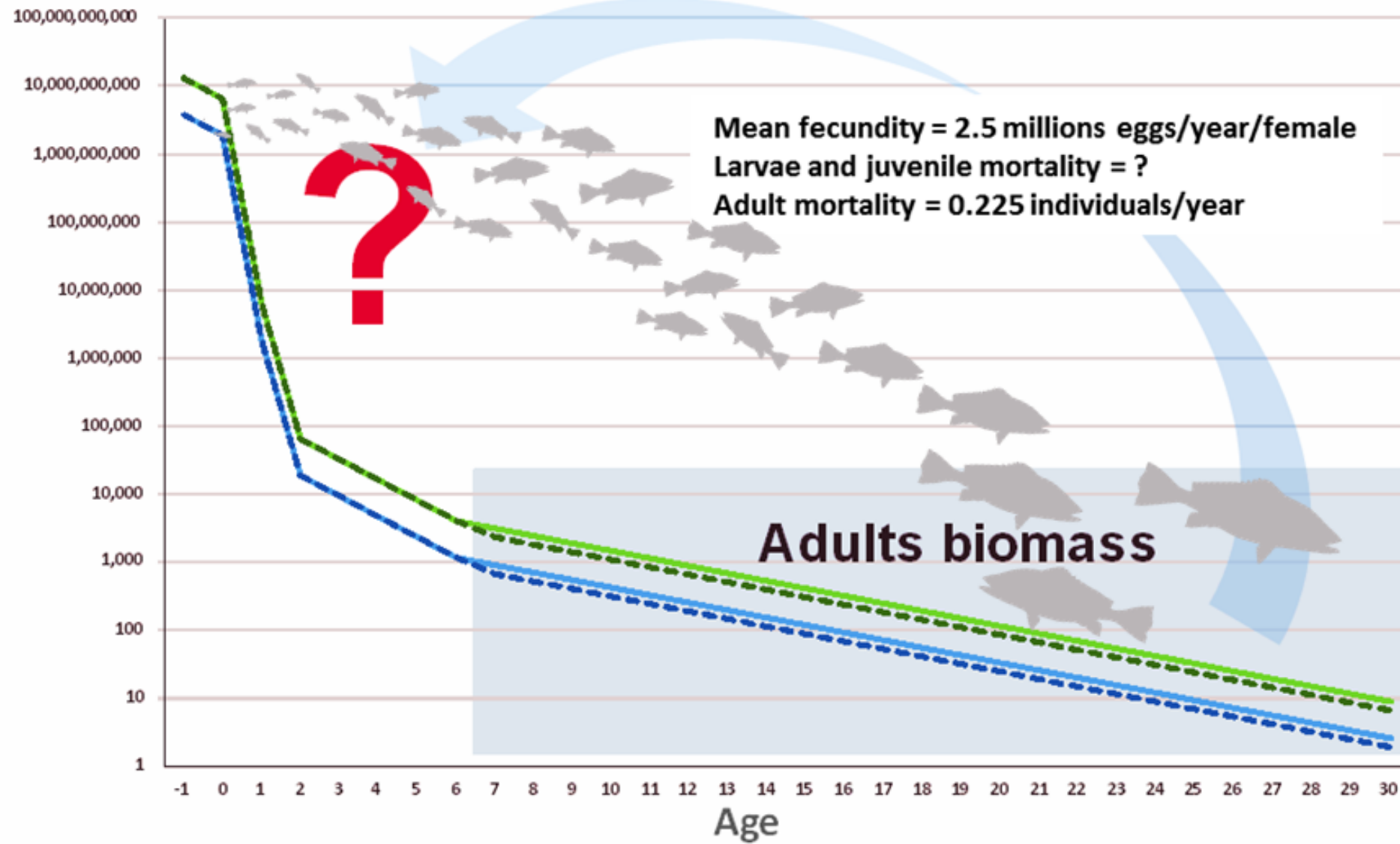
Markers	Method	n	N <sub>ef</sub>	N <sub>e</sub>	Reference
STRs (16)	MsVar	180	-	2,670	Valenzuela-Quiñones <i>et al.</i> 2014
STRs (16)	Nei	180	-	1,900	Valenzuela-Quiñones <i>et al.</i> 2014
STRs (16)	LD Ne Estimator	180	-	2,760	Valenzuela-Quiñones <i>et al.</i> 2014
STRs (16)	OneSamp	180	-	1,800	Valenzuela-Quiñones <i>et al.</i> 2014
STRs (16)	DIYABC	180	-	2,680	Valenzuela-Quiñones <i>et al.</i> 2014
STRs (24)	DIYABC	317	-	4,250	UABC 2015
mtDNA (1)	Temporal Method	573	5,370	10,740	UABC 2015
mtDNA (1)	DIYABC	792	5,560	11,120	UABC 2015
mtDNA (1)	BEAST-BSP	792	4,500	9,000	UABC 2015

## Census population size for 1 y.o. Cohort

Markers	Method	n	N	Reference
Multilocus genotype (2)	Capture-Recapture	119	150,000	UABC 2015



Individuals



Nef 5000  
251 MT

Nef 5000 Corr\*  
186 MT\*

Nef 1500  
72 MT

Nef 1500 Corr\*  
54 MT\*

\*Corrected for an estimated annual illegal catch of 50-70 MT